



# Standard Test Methods for Total and Dissolved Carbon Dioxide in Water<sup>1</sup>

This standard is issued under the fixed designation D 513; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon ( $\epsilon$ ) indicates an editorial change since the last revision or reapproval.

## 1. Scope\*

1.1 These test methods provide for the measurement of total or dissolved carbon dioxide present as carbon dioxide (CO<sub>2</sub>), carbonic acid, bicarbonate ion, and carbonate ion in water:

	Sections
Test Method A (Gas Sensing Electrode)	8-15
Test Method B (CO <sub>2</sub> Evolution, Coulometric Titration)	16-24

1.2 Carbon dioxide may also be detected from carbonates present in particulates in samples.

1.3 It is the user's responsibility to ensure the validity of these test methods on waters of untested matrices.

1.4 *This standard may involve hazardous materials, operations, and equipment. This standard does not purport to address all of the safety problems associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

1.5 Several test methods were discontinued from this standard in 1988. Refer to Appendix X1 for Historical Information.

## 2. Referenced Documents

### 2.1 ASTM Standards:

- D 1066 Practice for Sampling Steam<sup>2</sup>
- D 1129 Definitions of Terms Relating to Water<sup>2</sup>
- D 1192 Specification for Equipment for Sampling Water and Steam<sup>2</sup>
- D 1193 Specification for Reagent Water<sup>2</sup>
- D 1293 Test Methods for pH of Water<sup>2</sup>
- D 2777 Practice for Determination of Precision and Bias of Applicable Methods of Committee D-19 on Water<sup>2</sup>
- D 3370 Practices for Sampling Water<sup>2</sup>
- E 200 Practice for Preparation, Standardization, and Storage of Standard Solutions for Chemical Analysis<sup>2</sup>

## 3. Definitions

3.1 For definitions of terms used in these test methods, refer to Definitions D 1129.

## 4. Significance and Use

4.1 Carbon dioxide is a major respiration product of plants and animals and a decomposition product of organic matter and certain minerals. The atmosphere averages about 0.04 volume percent of CO<sub>2</sub>. Surface waters generally contain less than 10 mg/L, except at local points of abnormal organic or mineral decomposition; however, underground water, particularly deep waters, may contain several hundred mg/L.

4.2 When dissolved in water, CO<sub>2</sub> contributes significantly to corrosion of water-handling systems. This is particularly troublesome in steam condensate systems. Loss of CO<sub>2</sub> from an aqueous system can disturb the carbonate equilibrium and result in calcite encrustation of confining surfaces. Scaling of water heaters is a good example. Because of the delicate balance between corrosion and encrustation tendencies, much care must be given to control of CO<sub>2</sub> and related species in water systems. Recarbonation of municipal supplies during final stages of softening and amine neutralization of steam condensate are applied for these purposes.

## 5. Interferences

5.1 *General*—Carbon dioxide is easily lost from solution during transit and storage of samples. It also is possible for total CO<sub>2</sub> concentration to increase after sampling due to solution of finely divided calcium carbonate as a result of temperature or pressure changes.

## 6. Purity of Reagents

6.1 Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available.<sup>3</sup> Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

6.2 Unless otherwise indicated, references to water shall be understood to mean water conforming to Type I of Specification D 1193. Additionally, for those test methods

<sup>1</sup> These test methods are under the jurisdiction of ASTM Committee D-19 on Water and are the direct responsibility of Subcommittee D19.05 on Inorganic Constituents in Water.

Current edition approved Aug. 19, 1988. Published October 1988. Originally published as D 513 - 38. Last previous edition D 513 - 82.

<sup>2</sup> Annual Book of ASTM Standards, Vol 11.01.

<sup>3</sup> "Reagent Chemicals, American Chemical Society Specifications," Am. Chemical Soc., Washington DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see "Reagent Chemicals and Standards," by Joseph Rosin, D. Van Nostrand Co., Inc., New York, NY, and the "United States Pharmacopeia."

requiring water free of CO<sub>2</sub>, refer to Section 8.2 of Practice E 200.

## 7. Sampling

7.1 Collect the sample in accordance with Practice D 1066, Specification D 1192, and Practices D 3370, as applicable.

7.2 Filter samples when they are collected if particulates are present that may contain carbonates if dissolved species only are to be determined. When aliquots of sample are taken from sample bottles containing particulates, the bottle must be shaken or otherwise homogenized to ensure a representative sample is taken. When particulates form in samples due to changes in temperature, pH, etc., after the sample has been collected, these particulates must be included in tests of the sample. Care must be used to avoid loss of CO<sub>2</sub> during any homogenization or filtration of samples. Do not filter samples unless it is required to remove potentially interfering particulates.

7.3 Use a hard glass, chemically-resistant bottle for collecting the sample.

7.4 Fill the sample bottle completely, with no air space remaining beneath the cap, and store the sample at a temperature below that at which it was collected until the determination is made.

### TEST METHOD A—GAS SENSING ELECTRODE TEST METHOD

## 8. Scope

8.1 This test method determines total or dissolved carbon dioxide (9.2) present as CO<sub>2</sub>, carbonic acid, bicarbonate ion, and carbonate ion in water, within the interference constraints specified.

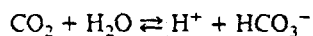
8.2 Samples containing 2 to 800 mL/L total CO<sub>2</sub> can be analyzed by this test method. The concentration range may be extended by dilution of an appropriate aliquot.

8.3 Samples should be analyzed immediately. If this is not possible, preserve by making them slightly alkaline (pH between 8 and 9) using carbonate-free NaOH solution and store them in a tightly capped vessel. The latter step prevents absorption of CO<sub>2</sub> from the air.

8.4 The precision and bias were obtained on reagent water and a water matrix of choice which included natural waters and brines. It is the responsibility of the analyst to determine the acceptability of this test method for the water being analyzed.

## 9. Summary of Test Method

9.1 Carbon dioxide is liberated by acidification of the sample to pH 5.0. The carbon dioxide electrode uses a gas-permeable membrane to separate the sample solution from the electrode internal solution. Dissolved carbon dioxide in the sample solution diffuses through the membrane until an equilibrium is reached between the partial pressure of CO<sub>2</sub> in the sample solution and the CO<sub>2</sub> in the internal filling solution. In any given sample, the partial pressure of CO<sub>2</sub> will be proportional to the concentration of CO<sub>2</sub>. The diffusion of CO<sub>2</sub> across the membrane affects the concentration of hydrogen ions in the internal filling solution:



The hydrogen ion concentration of the internal solution is measured by the pH electrode located behind the membrane. Since the hydrogen ion concentration is directly related to CO<sub>2</sub> concentration, the electrode response is Nernstian.

9.2 Samples are treated prior to measurement with a buffer solution which sets the pH at between 4.8 and 5.2. At this pH, interferences are minimized and the various ionic forms are converted to CO<sub>2</sub> (see Section 10).

## 10. Interferences

10.1 Volatile weak acids are potential positive electrode interferences. Concentrations of these interfering species that cause a 10 % error at 44 mg/L CO<sub>2</sub> or 100 mg/L (as CaCO<sub>3</sub>) and at pH 4 and 5, are listed below:

Interferences, mg/L	pH 5	pH 4
H <sub>2</sub> S	10	7
NO <sub>2</sub> <sup>-</sup> (NO <sub>2</sub> )	161	24
HSO <sub>3</sub> <sup>-</sup> (SO <sub>2</sub> )	320 (as SO <sub>2</sub> )	48 (as SO <sub>2</sub> )
HOAc (acetic acid)	372	216
HCOOH (formic acid)	1841	345

10.2 Samples containing sulfide can be treated with dilute solutions of potassium dichromate (or the like), since sulfur is not an interference for this test method. However, it is possible that some organic material could be oxidized to CO<sub>2</sub> by this treatment, resulting in falsely high results. The suitability of the test method for samples containing sulfide should be determined individually.

10.3 Water vapor is a potential electrode interference. Water can move across the membrane as water vapor, changing the concentration of the internal filling solution under the membrane. Such changes will be seen as electrode drift. Water vapor transport is not a problem if (1), the total concentration of dissolved species in solution (osmotic strength. Note 1) is approximately equal to that of the internal filling solution and (2); electrode and sample temperatures are the same.

NOTE 1—The osmotic strength of a solution is related to the total concentration of dissolved "species" in the solution. For example, the osmotic strength of a solution containing 0.1 M hydrochloric acid, 0.1 M acetic acid and 0.1 M sucrose is determined as follows: Hydrochloric acid dissociates to give 0.1 M hydrogen ion and 0.1 M chloride ion. The acetic acid, because of the concentration of free hydrogen ion, is essentially undissociated; thus giving 0.1 M of "species." Likewise, the concentration of sucrose "species" is 0.1 M. Therefore, the total osmotic strength is 0.4 osmolar.

10.4 Addition of carbon dioxide buffer (12.1) to samples of low-osmotic strength automatically adjusts them to the correct level. Samples with osmotic strength greater than approximately 1 M should be diluted before measurement, to avoid drifting associated with water vapor transport. Dilution should not reduce the carbon dioxide level below 2.5 mg/L. Samples with osmotic strengths above 1 M that cannot be diluted can be measured by adjusting the osmotic strength of the internal filling solution. To adjust the total concentration of dissolved species in the internal filling solution, add 0.425 g of reagent-grade NaNO<sub>3</sub> to 10 mL of internal filling solution.

## 11. Apparatus

11.1 pH Meter, with expanded mV scale, or a selective ion meter.

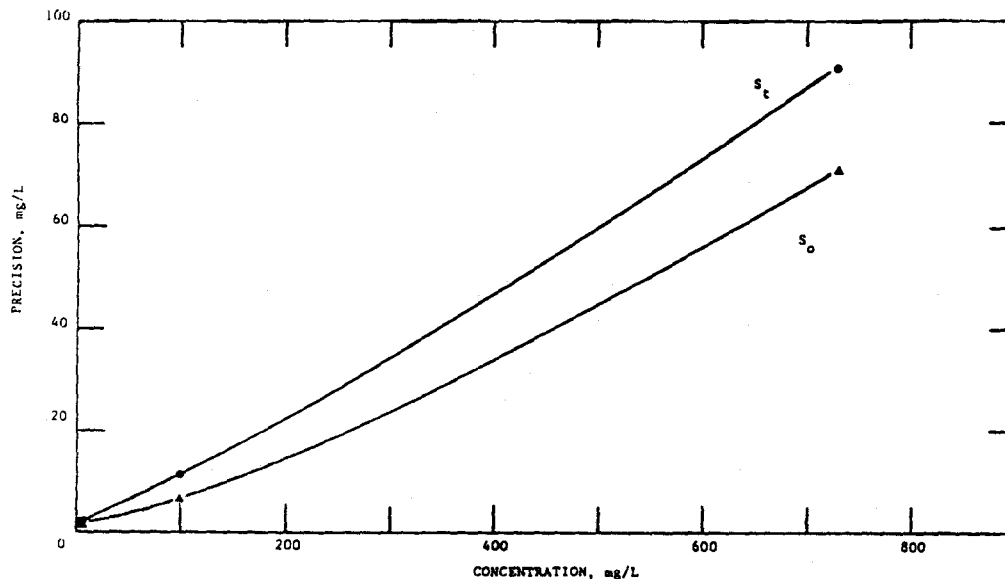


FIG. 1 Interlaboratory Precision for Total CO<sub>2</sub> Found in Reagent Water—Test Method A

11.2 CO<sub>2</sub> Gas-Sensing Electrode.<sup>4</sup>

11.3 Mixer, magnetic, with TFE-fluorocarbon-coated stirring bar, or equivalent.

12. Reagents

12.1 Buffer Solution—Dissolve 294 g of sodium citrate (Na<sub>3</sub>C<sub>6</sub>H<sub>5</sub>O<sub>7</sub>·2H<sub>2</sub>O) in approximately 700 mL of water in a 1-L volumetric flask. Acidify the solution to pH 4.5 with concentrated HCl (approximately 90 mL) and dilute to the mark with water.

12.2 Sodium Bicarbonate Solution, Standard (0.1 M)—Dissolve 8.40 g of sodium bicarbonate (NaHCO<sub>3</sub>) in water and dilute to 1 L.

12.3 Sodium Bicarbonate Solution, Standard (0.01 M)—Dilute 10.0 mL of sodium bicarbonate standard solution (0.1 M) to 100 mL.

13. Calibration

13.1 Assemble and check the electrode in accordance with the manufacturer's instructions.

13.2 Dilute 10 mL of the buffer solution to 100 mL with water using a volumetric flask. Transfer the contents of the flask to a 150-mL beaker and add a stirring bar. Immerse the electrode in the solution. Stir at a slow rate using the magnetic stirrer.

13.3 Using a volumetric pipet, add 0.5 mL of the 0.01 M NaHCO<sub>3</sub> standard solution and mix slowly. Allow the potential reading to stabilize (approximately 10 min) and record the potential (corresponds to 2.2 mg/L CO<sub>2</sub> or 5.0 mg/L (as CaCO<sub>3</sub>)).

13.4 Using a volumetric pipet, add 0.5 mL of the 0.01 M NaHCO<sub>3</sub> standard solution and mix slowly. Allow the potential reading to stabilize (approximately 5 min) and record the potential (corresponds to 4.4 mg/L CO<sub>2</sub> or 10.0 mg/L (as CaCO<sub>3</sub>)).

13.5 Using a volumetric pipet, add 0.9 mL of the 0.1 M NaHCO<sub>3</sub> standard solution and mix slowly. Allow the potential reading to stabilize (approximately 2 min) and record the potential (corresponds to 43.2 mg/L CO<sub>2</sub> or 98.1 mg/L (as CaCO<sub>3</sub>)).

13.6 Using a volumetric pipet, add 10 mL of the 0.1 M NaHCO<sub>3</sub> standard solution and mix slowly. Allow the potential reading to stabilize (approximately 2 min) and record the potential (corresponds to 433 mg/L CO<sub>2</sub> or 983 mg/L (as CaCO<sub>3</sub>)).

13.7 Plot potential values (on the linear scale) versus concentration (on the logarithmic scale) on semilogarithmic graph paper to obtain a calibration curve. The curve may be extended down to 2 mg/L and up to 800 mg/L CO<sub>2</sub>.

14. Procedure

14.1 Bring samples to same temperature as the electrode and standards.

14.2 Place a known volume, V<sub>s</sub> (100 mL is convenient) of sample in 150-mL beaker and stir slowly. Immerse the electrode in the solution.

14.3 Add 1 mL of buffer, V<sub>b</sub>, for each 10 mL of sample. Allow the potential reading to stabilize and record the value. Read the concentration measured (C<sub>m</sub>) directly from the calibration curve.

14.4 Determine the sample concentration (C<sub>s</sub>) as follows:

$$C_s = C_m \frac{V_s + V_b}{V_s}$$

15. Precision and Bias<sup>5</sup>

15.1 Precision—The overall and single-operator precision of this test method, within its designated range, varies with the quantity tested as shown in Fig. 1, for reagent water, and Fig. 2, for selected water matrices. These matrices included natural waters and brines.

<sup>4</sup> Orion Model 95-02, or equivalent, available from Orion Research Inc., 529 Main St., Boston, MA 02129.

<sup>5</sup> Supporting data for this test method are available from ASTM Headquarters. Request RR:D19-1069.

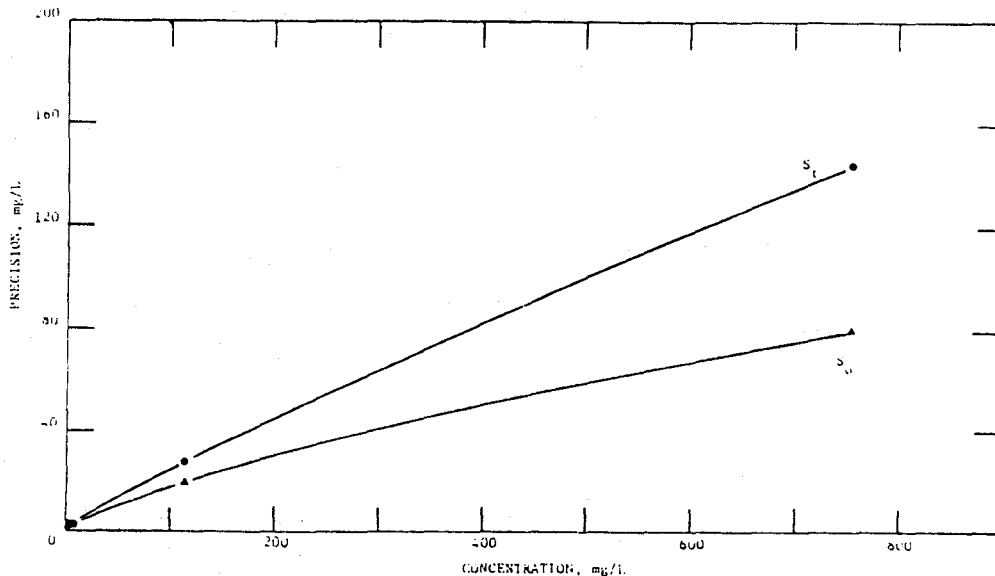


FIG. 2 Interlaboratory Precision for Total CO<sub>2</sub> Found in Selected Water Matrices—Test Method A

TABLE 1 Bias—Test Method A

Amount Added, mg/L	Amount Found, mg/L	± % Bias	Statistically Significant (91 % Confidence Level)
Reagent water			
2.1	2.7	+28.6	no
5.1	6.9	+35.3	yes
101	100	-1.0	no
803	730	-9.1	yes
Water matrices			
2.1	3.0	+42.8	no
5.1	7.1	+39.2	yes
101	114	+12.9	no
803	754	-6.1	no

15.2 *Bias*—Recoveries of known amounts of total CO<sub>2</sub> from reagent water and selected water matrices were as shown in Table 1.

15.3 The information in 15.1 and 15.2 is derived from round-robin testing in which eight laboratories, including twelve independent operators, participated. Of twelve data sets ranked as described in Practice D 2777, four were rejected in the case of reagent water and three were rejected in the case of selected water matrices. Four “outlier” data points were also rejected. Four sample levels were run on three days, and blanks were obtained for the waters used.

TEST METHOD B—CO<sub>2</sub> EVOLUTION, COULOMETRIC TITRATION TEST METHOD

16. Scope

16.1 This test method determines total or dissolved carbon dioxide present as carbon dioxide (CO<sub>2</sub>), carbonic acid, bicarbonate ion, and carbonate ion in water within the interference constraints specified.

16.2 Carbon dioxide will also be detected from carbonates present in particulates in samples.

16.3 Samples containing between 5 and 800 mg/L total CO<sub>2</sub> can be analyzed by this test method. The concentration range may be extended upward by use of smaller samples or

dilution of an appropriate aliquot. The range may be extended lower by use of larger samples.

16.4 The precision and bias information reported in this test method was obtained in collaborative testing which included waters of the following types: distilled, deionized, potable, natural, brine, industrial waste, and waters derived from oil shale retorting. Since the precision and bias information reported may not apply to waters of all matrices, it is the user’s responsibility to ensure the validity of the test method on samples of other matrices.

17. Summary of Test Method

17.1 Carbon dioxide is liberated by acidifying and heating the samples. The liberated CO<sub>2</sub> is swept through a scrubber by carbon dioxide-free air into an absorption cell where it is automatically coulometrically titrated. Concentrations of the several carbonate species are determined from the pH and total CO<sub>2</sub> values.

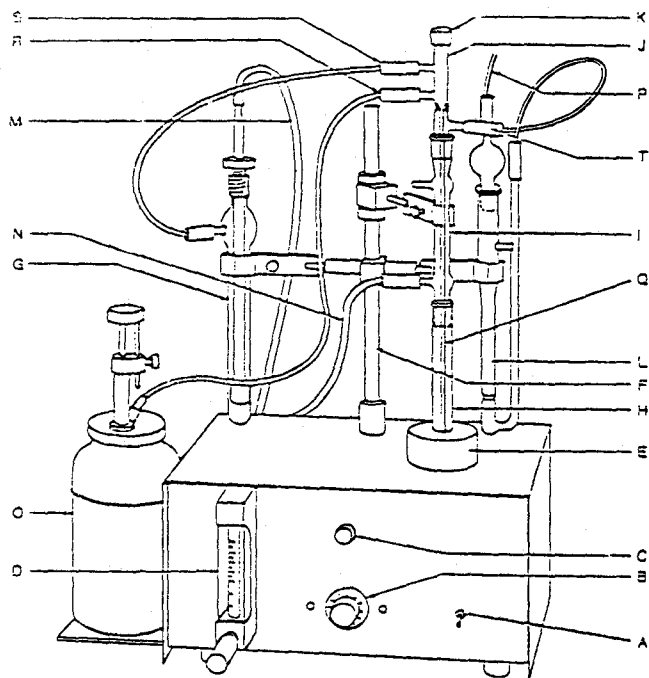
18. Interferences

18.1 Any volatile acid or base not removed by the scrubbing solution will interfere with the test. Potentially interfering gases which the scrubber removes include: hydrogen sulfide (H<sub>2</sub>S), chlorine (Cl<sub>2</sub>), bromine (Br<sub>2</sub>), hydrogen chloride (HCl), hydrogen bromide (HBr), hydrogen iodide (HI), hydrogen fluoride (HF), sulfur dioxide (SO<sub>2</sub>), and sulfur trioxide (SO<sub>3</sub>).

18.2 If a significant precipitate forms in the scrubber solution during an analysis, the scrubber solution should be replaced and the analysis repeated.

18.3 If the level of potentially interfering materials is such that the scrubber capacity is exhausted rapidly, an additional higher capacity scrubber may be added as directed under 19.3. When two scrubbers are used the scrubber capacity is considered to be exhausted when a precipitate begins forming in the final scrubber.

18.4 When analyzing samples that may evolve acid gases other than those listed, previously spiked samples should be



- |                        |                                 |
|------------------------|---------------------------------|
| A Main power           | K Septum                        |
| B Heater control       | L Sample scrubber               |
| C Heat indicator light | M Analysis air line             |
| D Flow meter           | N Condenser air line            |
| E Heater and shield    | O Acid dispenser                |
| F Main support         | P Scrubber outlet to coulometer |
| G Air scrubber         | Q Sample purge tube             |
| H Sample tube          | R Acid inlet                    |
| I Condenser            | S Purified air inlet            |
| J Adaptor tube         | T Sample air outlet             |

FIG. 3 Carbonate Carbon (CO<sub>2</sub>) Apparatus—Test Method B

analyzed to confirm that the scrubber(s) used are effective and scrubbers should be modified if necessary.

## 19. Apparatus

19.1 *Carbon Dioxide Coulometer* to automatically titrate evolved carbon dioxide.<sup>6</sup>

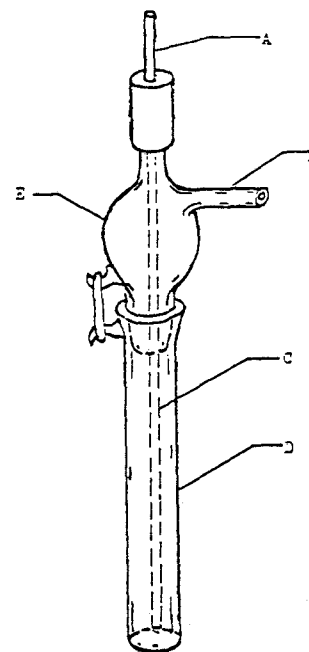
19.2 *Carbon Dioxide Evolution Apparatus*, consisting of an air pump, air purification scrubber, acid dispenser, sample reaction chamber, and scrubber. The arrangement of the apparatus is shown in Fig. 3.<sup>7</sup>

19.3 *High-Capacity Scrubber*, if needed, may be assembled as illustrated in Fig. 4. It should be installed so that the gas flows from the sample air outlet through the scrubber into the regular scrubber. The size may be increased over that specified, but doing so may lengthen the analysis time.

19.4 *pH Meter*, conforming to the requirements given in Test Method D 1293.

## 20. Reagents

20.1 *Coulometer Cell Reagents*—Cell solutions to absorb CO<sub>2</sub> from the gas stream and convert it to a titratable acid



- |   |
|---|
| A Inlet, from sample air outlet   |
| B Outlet, to standard scrubber  |
| C Tube, extends into scrubber solution  |
| D Body, glass with 1/2 standard-taper joint, total length 125 mm, used with up to 12-mL scrubber solution |
| E Adaptor, glass  |

FIG. 4 High-Capacity Scrubber—Test Method B

and permit 100 % efficient coulometric titration.<sup>7</sup>

20.2 *Perchloric Acid Solution* (approximately 1+5)—Add 170 mL of concentrated HClO<sub>4</sub> (sp gr 1.67) to 500 mL of water, mix, and dilute to 1 L with water.

20.3 *Potassium Hydroxide Solution* (65 g/100 mL)—Dissolve 65 g of KOH in water and dilute to 100 mL with water.

20.4 *Scrubber Solution*—Add 2 mL of concentrated HClO<sub>4</sub> (sp gr 1.67) and 5 g of silver percholate (AgClO<sub>4</sub>) to 100 mL of water. When adding scrubber solution to the scrubber(s), and 0.1 mL of 30 % hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) to the scrubber per mL of scrubber solution used in the scrubber. As an alternative scrubber solution, use 2 mL of concentrated sulfuric acid (H<sub>2</sub>SO<sub>4</sub>) per 100 mL of water saturated with silver sulfate (Ag<sub>2</sub>SO<sub>4</sub>) and add 30 % of H<sub>2</sub>O<sub>2</sub> as for the AgClO<sub>4</sub> scrubber solution.

## 21. Calibration

21.1 Calibration is not required; however, standards should be analyzed before analyzing samples and periodically to confirm proper operation of the instrument. If the recovery of a standard is unacceptable, the cause of the poor result should be determined and corrected. Generally low results are due to leaks and high results due to contamination of reagents or an exhausted scrubber(s).

## 22. Procedure

### 22.1 Blank Determination:

22.1.1 Set up the CO<sub>2</sub> coulometer as directed by the instrument manufacturer.

<sup>6</sup> Instrument marketed by Coulometrics, Inc., a subsidiary of UIC Inc., P.O. 863, Joliet, IL 60434.

<sup>7</sup> Satisfactory reagents available from Coulometrics, Inc. use ethanolamine to absorb CO<sub>2</sub> forming hydroxyethylcarbamic acid which is titrated coulometrically using a color indicator for end-point detection.

22.1.2 Acidify 100 mL of water to pH between 2 and 3 with HClO<sub>4</sub> (1+5) and boil vigorously for at least 15 min to remove dissolved CO<sub>2</sub>.

22.1.3 Inject 5.00 mL of selected sample size (see 22.2) of the freshly boiled water into the apparatus using a calibrated syringe. Set the coulometer display to 0, pump 2 mL of HClO<sub>4</sub> (1+5) into the reaction tube, and position the reaction tube over the heater. After 5 min record the coulometer displays as *B*.

22.1.4 Make a blank determination before each series of CO<sub>2</sub> determinations.

22.1.5 Empty the reaction tube following completion of an analysis if the remaining volume is insufficient for the next sample plus acid. The maximum liquid volume of the sample tube is 12 mL. When the sample tube is replaced, CO<sub>2</sub> from air which entered the apparatus must be swept out by the gas stream before beginning the next analysis. Sixty seconds is normally sufficient for this. To eliminate the necessity of removing the sample tube and the subsequent purge time, a stopcock may be attached to the sample tube to permit draining it following an analysis.

22.2 Carbon Dioxide Determination:

22.2.1 For samples with less than 800 mg/L of CO<sub>2</sub>, use the procedure described in 22.1.3 for blank determinations on a 5.00-mL untreated portion of the sample. Record the coulometer readout as *A*.

22.2.2 If the CO<sub>2</sub> content of the sample exceeds 800 mg/L, use a proportionately smaller sample and adjust the calculations accordingly. Do not reduce sample volumes so much that representative samples cannot be obtained or sample volumes cannot be measured with sufficient accuracy. Alternatively, the sample may be diluted using CO<sub>2</sub>-free water. For samples containing less than 100 mg/L, accuracy may be increased by use of a larger sample size if a larger sample tube is used.

22.2.3 If the volume of the sample tube or scrubbers is increased, the analysis time may have to be lengthened. At the end of an analysis the titration should be complete and the coulometer display stable. If the titration is not complete, the analysis time must be lengthened for both blanks and samples.

22.2.4 If desired, samples may be pipeted into acid free reaction tubes positioned on the apparatus to begin the analysis. In this case, sufficient time (generally 60 s) must be allowed for CO<sub>2</sub> from air that entered the apparatus to be swept from the apparatus before acid is added and the analysis begun.

NOTE 2—For some samples a significant amount of CO<sub>2</sub> may be removed from the sample during the purge of the apparatus. This can be determined by comparing the amount of CO<sub>2</sub> removed during the purge of a blank and the purge of the sample.

22.2.5 If 2 mL of HClO<sub>4</sub> (1+5) is not sufficient to acidify the samples to pH 3 or less, more acid may be used or more concentrated HClO<sub>4</sub> used. Sulfuric acid (H<sub>2</sub>SO<sub>4</sub>) may be substituted for HClO<sub>4</sub>. When samples include sediments or other particulate matter, wetting and emulsifying agents may

be added to acid to ensure more rapid reaction between the solids and the acid.

22.3 Determine the pH of the original sample in accordance with Test Method D 1293.

NOTE 3—If the temperature of the sample at the time of the test is different from its temperature at the time of collection, a different CO<sub>2</sub>-HCO<sub>3</sub><sup>-</sup>-CO<sub>3</sub><sup>=</sup> equilibrium and a different pH will result. This may be partially overcome by adjusting the sample to the collection temperature and shaking it thoroughly before opening it for CO<sub>2</sub> and pH tests. It is best to avoid the problem by measuring pH at the time of sample collection and sealing the sample to prevent loss or gain of CO<sub>2</sub>.

23. Calculation

23.1 Calculate the concentration of the total CO<sub>2</sub> using 23.1.1 or 23.1.2 and 23.1.3 as appropriate for the coulometer display units and the sample volume used.

23.1.1 CO<sub>2</sub> coulometer set to display micrograms of carbon:

$$\text{Total CO}_2, \text{ mg/L} = 0.7329 (A - B)$$

where:

*A* = carbon from sample, μg, and  
*B* = carbon from blank determination, μg.

23.1.2 CO<sub>2</sub> coulometer set to display micrograms of carbon dioxide:

$$\text{Total CO}_2, \text{ mg/L} = 0.200 (A - B)$$

where:

*A* = carbon dioxide from sample, μg, and  
*B* = carbon dioxide from blank determination, μg.

23.1.3 If the sample size used is not 5.00 mL, adjust the calculation accordingly by multiplying the value calculated in 23.1.1 or 23.1.2 by 5/mL sample volume.

24. Precision and Bias<sup>5</sup>

24.1 Precision—The single-operator and overall precision of this test method within its designated range varies with the quantity being tested according to Fig. 5.

24.2 Bias—Recoveries of known amounts of CO<sub>2</sub> in a series of spiked samples were:

Amount added mg/L	Amount recovered mg/L	± Bias	± % Bias	Significant
7.8	8.09	+0.29	+3.72	No
22.1	22.29	+0.19	+0.86	No
372.1	369.83	-2.37	-0.64	Yes
736.0	730.87	-5.13	-0.70	Yes

24.3 The precision and bias data were derived from results of the cooperative tests on samples prepared by laboratories spiking a matrix(s) of choice with solutions of NaHCO<sub>3</sub>. Eight laboratories participated. Samples of interest chosen by participating laboratories included distilled water, deionized water, potable water, natural waters, brine, industrial waters, and waters derived from oil shale retorting. The procedure may not apply to all matrices. It is the user's responsibility to ensure the validity of this test method on other sample matrices and to determine the effect on the precision and bias when other than 5.00-mL sample volumes are used.

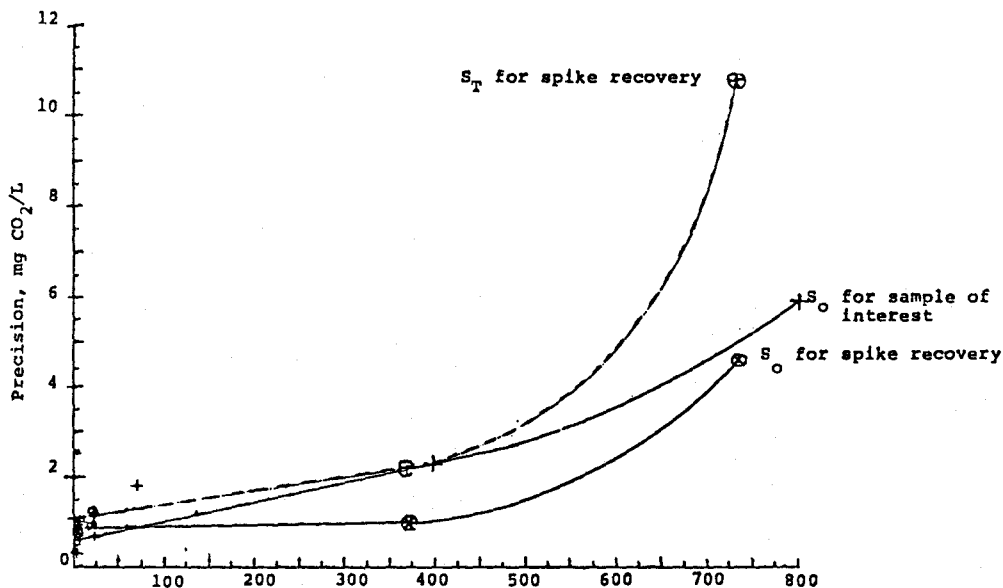


FIG. 5 Interlaboratory Precision for CO<sub>2</sub> Found in Water by Acid Evolution and Coulometric Titration—Test Method B

## APPENDIX

### (Nonmandatory Information)

#### X1. RATIONALE FOR DISCONTINUATION OF TEST METHODS

X1.1 The test methods briefly described in X1.1.1 through X1.1.5 were discontinued in 1988. They are published in their entirety in the 1987 *Annual Book of ASTM Standards*, Vol 11.01.

X1.1.1 *Precise CO<sub>2</sub> Evolution Test Method*—Carbon dioxide is liberated by acidifying and heating the sample in a closed system, which includes a condenser, a gas scrubber, a CO<sub>2</sub> absorber, an expansion bladder, and a gas-circulating pump. The liberated CO<sub>2</sub> is combined with barium hydroxide in an absorber, and the excess hydroxide is titrated with standard acid. Concentrations of the several carbonate species are determined from the pH and total CO<sub>2</sub> values.

X1.1.2 *Abridged CO<sub>2</sub> Evolution Test Method*—Carbon dioxide and bicarbonate ion are fixed with sodium hydroxide and precipitated as strontium carbonate. The solution is then neutralized, the CO<sub>2</sub> is removed by aeration in the presence of excess acid, and the quantity removed is determined by back-titration of the acid.

X1.1.3 *Bicarbonate Titration Test Method*—Carbon dioxide concentration is determined from measured values of pH and bicarbonate ion.

X1.1.4 *Differential Titration Test Method*—The water sample is titrated to pH 8.5 and pH 5.0 using standard alkali and acid as appropriate. It then is acidified, boiled to remove CO<sub>2</sub>, retitrated to the same two pH points, and the total CO<sub>2</sub> content is calculated.

X1.1.5 *Direct Titration of Free CO<sub>2</sub>*—Free CO<sub>2</sub> is reacted with sodium hydroxide to form sodium bicarbonate. The end point of the reaction is detected electrometrically or by means of a pH color indicator.

X1.2 The test methods in X1.1.1 through X1.1.5 were discontinued because there were insufficient laboratories interested in participating in the collaborative studies needed to provide the precision and bias data required by Practice D 2777.

## SUMMARY OF CHANGES

This section identifies the location of selected changes to these test methods that have been incorporated since the last issue. For the convenience of the user, Committee D-19 has highlighted those changes that may impact the use of these test methods. This section may also include descriptions of the changes or reasons for the changes, or both.

(1) Test Methods A (Precise CO<sub>2</sub> Evolution), B (Abridged CO<sub>2</sub> Evolution), C (Bicarbonate Titration), D (Differential Titration), and E (Direct Titration of Free CO<sub>2</sub>) were discontinued because there were insufficient laboratories interested in participating in the collaborative studies needed to provide the precision and bias data required by Practice D 2777.

*The American Society for Testing and Materials takes no position respecting the validity of any patent rights asserted in connection with any item mentioned in this standard. Users of this standard are expressly advised that determination of the validity of any such patent rights, and the risk of infringement of such rights, are entirely their own responsibility.*

*This standard is subject to revision at any time by the responsible technical committee and must be reviewed every five years and if not revised, either reapproved or withdrawn. Your comments are invited either for revision of this standard or for additional standards and should be addressed to ASTM Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee, which you may attend. If you feel that your comments have not received a fair hearing you should make your views known to the ASTM Committee on Standards, 1916 Race St., Philadelphia, PA 19103.*